
Review

The Interleukin-21 (IL-21) Signal and Allergic Immune Response

Osam Mazda* and Tsunao Kishida

*Department of Microbiology and Immunology,
Kyoto Prefectural University of Medicine Graduate School of Medical Science **

Abstract: Interleukin-21 (IL-21) is a member of the common γ (γ c) chain family of cytokines, mainly produced by Th17 cells and natural killer T (NKT) cells. It plays pleiotropic immunomodulatory roles through acting on various target cells including T cells, B cells, NK cells, macrophages and dendritic cells. IL-21 may be involved in pathogenesis of some autoimmune and malignant diseases. IL-21 signal counteracts IL-4 and inhibits IgE class switch recombination (CSR) in B cells, in which the inhibitor of differentiation 2 (Id2) is induced by IL-21, and essentially contributes to the IgE CSR regulation. Exogenous IL-21 drastically suppresses anaphylactic and local allergic reactions in animals. Molecular understanding of the IL-21-mediated suppression of allergic responses may provide important information for development of novel diagnostic and therapeutic modalities to control allergic diseases.

Key Words: Interleukin-21, Id2, IgE, Class switch, Allergy.

The IL-21 and IL-21 signal

IL-21 is a four- α -helix bundle cytokine with a significant amino acid sequence homology to IL-2, IL-4, and IL-15¹⁾. It exerts a variety of immunomodulatory functions through interaction with the cell surface IL-21 receptor (IL-21R) complex that is composed of two polypeptide chains, the IL-21R and common γ (γ c) chain. The former has sequence homology with the β component of the IL-2R complex and possesses the WSXWS motif. IL-21 binds to the IL-21R even in the absence of γ c chain, but IL-21R alone is not capable of transmitting a signal. The γ c chain is an essential signal transducing subunit shared by the receptor complexes for IL-2, IL-4, IL-7, IL-9 and IL-15¹⁾. Binding of IL-21 to the IL-21R/ γ c complex leads to phosphorylation of the Janus kinase (Jak) 1 and Jak 3, and the phosphorylation of the IL-21R at tyrosine 510, subsequently leading to the recruitment and phosphorylation of the signal transducer and activator of transcription 3 (STAT3)²⁾. The activation of STAT3 is essential for the IL-21 signaling²⁾. STAT4 and STAT5 are also phosphorylated upon IL-21 stimulation, but their activation are relatively weak and transient, in sharp contrast to IL-2 and IL-15 signals that strongly activate

STAT5 but not STAT3. Moreover, Ras-Raf-MAPK and PI3K pathways are also activated by the engagement of the IL-21R complex³.

IL-21 is produced by activated CD4⁺ T cells, including Th1, Th2, Th17 and T follicular helper cells⁴. Among them, Th17 cells most notably produces IL-21, which subsequently acts on Th17 in an autocrine fashion⁵⁻⁷. Meanwhile, natural killer T (NKT) cells are another producer of this cytokine⁸. CD8⁺ T cells also produce IL-21 under certain conditions⁹.

Parrish-Novak et al. reported that IL-21 cooperates with IL-2 or IL-15 to enhances NK cells, while IL-21 provokes differentiation of bone marrow NK cell progenitors in cooperation with IL-15 or flt3 ligand¹⁰. IL-21 also enhances proliferation of mature B and T cells, when combined with CD40 and CD3 antibodies, respectively.

IL-21 signal may play a crucial role in the commitment of naïve CD4⁺ T cells to Th17 differentiation. IL-21 provokes TGF- β - and IL-6-stimulated CD4⁺ T cells to produce IL-21, while inducing expression of ROR γ t, the Th17-specific transcriptional factor, and IL-23R⁵⁻⁷. Acting on naïve CD4⁺ T cells, IL-21 suppresses expression of STAT4, attenuates susceptibility to IL-12, and suppresses production of interferon- γ (IFN- γ)^{11,12}, so that Th1 differentiation may be inhibited. IL-21 also prevents TGF- β -mediated induction of regulatory T (Treg) cells.

IL-21 also exerts regulatory functions on NK cells, DCs and macrophages. IL-21 signal facilitates induction of cytotoxic T lymphocytes (CTL)¹³, and synergizes with IL-7 or IL-15 to enhance proliferation of CD8⁺ T cells¹⁴. IL-21 up-regulates expression of IL-4R α and IL-13R α 1 in macrophages, while inducing expression of CXCL8 that promotes migration of neutrophils¹⁵. IL-21 suppresses differentiation of murine bone marrow-derived DCs in vitro, whereas IL-21-stimulated DCs no longer provoke T cells to exert contact hypersensitivity reaction in vivo¹⁶. When human monocyte-derived DCs are stimulated with IL-21, SOCS-1 and SOCS-2 expression is elevated, while toll-like receptor (TLR) signal is attenuated. Moreover, IL-21 suppresses lipopolysaccharide (LPS)-induced production of TNF- α , IL-21, CCL5 and CXCL10 in the human monocyte-derived DCs¹⁷.

IL-21 exerts variety of actions on B cells. IL-21 may either positively or negatively regulate proliferation of B cells depending on the presence or absence of CD40¹⁸⁻²⁰, BCR^{18,21}, LPS or CpG DNA^{18,20} signals. In the presence of BCR and CD40 stimuli, IL-21 provokes naïve and memory B cells to differentiate into postswitch antigen-secreting cells (plasma cells)^{22,23}. IL-21 augments proliferation of B cells that has received BCR and CD40 signals, whereas IL-21 suppresses growth of, and induces apoptosis in, B cells that has received TLR-4 or 9 signal^{18,24}. This apoptosis is Bcl-1-sensitive, dependent on the caspase pathway, and mediated through induction of Bim, Bcl-6 and JunK, as well as through down-regulation of Bcl-xL^{18,24}. IL-21 up-regulates expression of Blimp-1 and facilitates terminal differentiation of B cells to antibody producing cells^{25,26}. Recently IL-21R signal was found critical for the development of memory B cell response²⁷.

IL-21 and diseases

Reportedly, the IL-21 signal was involved in the regulation of growth, survival and activation of lymphoma cells, while it promoted proliferation of acute T cell leukemia and multiple myeloma cells²⁸. Meanwhile, IL-21 induced apoptosis in B cell chronic lymphoma cell. Acting on the Epstein-Barr virus (EBV)-transformed B cell lines, IL-21 transiently enhanced proliferation, followed by subsequent induction of growth arrest and apoptosis, while expression of Granzyme B was up-regulated by IL-21

in EBV-transformed B cells^{9/28}).

We previously reported *in vivo* action of IL-21 on anti-tumor immunity. By transfecting mouse IL-21 (mIL-21) gene into mice bearing preestablished lung metastasis of RLmale1 lymphoma, we showed that IL-21 induced both CTL and NK cytolytic activities²⁹. We also demonstrated, using a head and neck squamous cell carcinoma model, that IL-21 elicited not only cellular immune responses mediated by CTL and NK cells but also humoral immune responses in which tumor-specific IgG antibody was produced and partially contributed to tumor rejection³⁰.

It has been suggested that IL-21 signal may be involved in the pathogenesis and progression of some autoimmune, including systemic lupus erythematosus, rheumatoid arthritis, multiple sclerosis^{31/32}. IL-21 may also be involved in some skin diseases such as psoriasis^{9/33} and infectious diseases. For an example, IL-21 may play important roles in anti-virus immune response, by affecting primary and memory anti-virus CD8⁺ T cell responses³⁴.

IL-21 and allergic reactions

Recent studies on human genome epidemiology suggested that the IL-21 signal may influence IgE production and be involved in allergic phenotype. Hecker et al. reported significant correlation between a single nucleotide polymorphism (SNP) of the IL-21R gene locus, i.e., T-83C, and elevated IgE levels in healthy female volunteer³⁵. Chatterjee et al. found that the C5250T polymorphism was significantly associated with atopic asthma and serum total IgE level³⁶.

There are some inconsistent reports on the influence of IL-21 on generation of IgE-producing B cells. Mice genetically deficient for IL-21R²⁶ or IL-21³⁷ exhibited more abundant production of IgE than did wild type mice in response to immunization with antigens. IL-21 reduced serum IgE levels in antigen-primed mice. Meanwhile, the cytokine suppressed IgE production in murine splenic B cells activated *in vitro* by LPS plus IL-4 due to the inhibition of IgE CSR²⁵. Another report demonstrated that IL-21 enhanced production of IgE by unfractionated peripheral blood mononuclear cells (PBMC) or B cells cultured in the presence of CD40 antibody and either IL-4 or IL-13³⁸. In this report, however, IL-21 suppressed the IgE production by PBMC stimulated by PHA and IL-4³⁸. Another report showed different notion that IL-21 rather elevated IgE production and proliferation of IgE producing B cells when the cytokine was added to PBMC or tonsillar B cells stimulated by CD40 antibody and IL-4³⁹. Also, when murine B cells was cultured with CD40 antibody and IL-4, IgE production was either increased or decreased by additional IL-21 depending on the cell density in the culture³⁹. It was also reported that IL-21 failed to inhibit IL-4-induced IgE CSR in human B cells derived from peripheral blood or spleen, but down-regulated IgE production through induction of IFN- γ ⁴⁰. The IFN- γ -mediated inhibition of IgE synthesis by IL-21 was associated with a polymorphism in the IL-21R gene⁴⁰.

IL-21 signal counteracts the IL-4 signal and abolishes IgE CSR in B cells

As described above, it remained controversial whether IL-21 directly acts on B cells and suppresses IgE CSR, while molecular mechanisms underlying the connection between IL-21 signal and IgE CSR had not been clarified. So we examined *in vitro* action of IL-21 on B cells that were provoked to undergo CSR. Splenic B cells from AKR/J mice were stimulated with IL-4 and LPS, so that significant amount of various classes of immunoglobulin was produced. An addition of recombinant mouse IL-21 (rmIL-21) significantly, and specifically, inhibited the production of IgE, clearly showing that IL-21

canceled the effect of rmIL-4 and LPS that otherwise induced IgE producing B cells⁴¹.

To figure out the molecular basis of the IgE suppression, splenic B cells derived from normal mice were cultured in the presence or absence of LPS, rmIL-4 and rmIL-21, and RT-PCR analysis was performed to detect germ line C ϵ transcript. An addition of both rmIL-4 and LPS significantly up-regulated germ line C ϵ transcript, indicating that these stimuli provoked B cells to undergo IgE CSR. In contrast, an addition of rmIL-21 almost completely abrogated the germ line C ϵ transcript, suggesting effective obstruction of IgE CSR by rmIL-21 that counteracted IL-4 and LPS signal⁴¹.

The IgE CSR is a critical step in IgE-based allergic reaction. Although the mechanism of CSR has not been completely understood, it has been widely accepted that the germ line Ig heavy chain gene transcript and the activation-induced cytidine deaminase (AID) plays pivotal roles for the CSR⁴²⁻⁴⁵. Three transcriptional factors, Paired box protein 5 (Pax5)⁴⁶, E2A⁴⁷ and STAT6^{48/49} positively regulate both the AID and germ line C ϵ transcription. Pax5 is constitutively expressed in B cells except for terminal differentiated plasma cells, and further up-regulated by CD40-mediated signal⁵⁰. E2A is induced by LPS, IL-4 plus anti-IgM, or IL-4 plus CD40⁵¹, while STAT6 is activated by either IL-4 or IL-13 signal⁵².

We further analyzed which molecule is involved in the IL-21-mediated suppression of these pathways. It was demonstrated that the inhibitor of differentiation 2 (Id2) is transcriptionally induced by IL-21 and the Id2 was essential and sufficient for the IL-21-mediated abrogation of IgE CSR⁴¹. Id proteins contain the helix-loop-helix (HLH) motif but lack a DNA binding domain^{53/54}. They antagonize the transactivation functions of the E proteins, which otherwise regulate many aspects of hematopoiesis as well as lymphocyte proliferation and survival through modulating gene expression program^{53/55-57}. It was found that the balance between Id2 and Pax5 is pivotal for the regulation of AID gene expression⁵⁸ and that Id2 negatively regulate IgE CSR⁵⁹.

Exogenous IL-21 suppresses systemic and local allergic responses

We examined effect of mIL-21 gene transfer on experimental peanut anaphylaxis that had been established by repetitive priming with crude peanut extract (CPE) as an allergen in combination with cholera toxin and aluminum hydroxide. After intragastric challenge of CPE anaphylactic symptoms were estimated by measuring rectal temperature as well as scoring the respiratory and peripheral circulatory disturbance, neurological manifestations, and survival of the animals. The signs of anaphylaxis were totally alleviated by intravenous transfection with the mIL-21 gene, while the untreated allergic mice showed severe systemic manifestation of anaphylactic reactions. The serum IgE level in the IL-21 gene-treated allergic animals was reduced to virtually the same level as that in normal mice. The mIL-21 gene treatment drastically decreased serum level IgE, while other classes of immunoglobulin were not significantly affected. Similar results were also obtained when rmIL-21 was injected intraperitoneally to CPE-primed mice instead of mIL-21 gene transfection.

To assess effect of IL-21 on IgE CSR, germ line C ϵ transcript was examined. It was clearly demonstrated that the rmIL-21 treatment drastically inhibited IgE CSR that was otherwise induced in B cell after CPE immunization. This may, at least partly, contributed to the IL-21-mediated decrease of IgE and amelioration of anaphylaxis. We also found that Id2 also plays an essential role in the IL-21-mediated inhibition of IgE CSR *in vivo*⁴¹.

We also tested whether IL-21 is capable of affecting local allergic responses. Allergic rhinitis model

of mice was established by repetitively administrating ovalbumin as allergen in combination with an adjuvant. Intra-nasal administration of rmIL-21 partially, but significantly, reduced allergic rhinitis symptoms. The OVA-specific IgE was reduced in the sera, and the germ line C ϵ transcript was drastically inhibited in the nasal-associated lymphoid tissues (NALT) of IL-21-treated mice⁶⁰.

As the molecular mechanisms of the IL-21-mediated suppression of IgE CSR, we found that Bcl-6 that is involved in germ line C ϵ transcription is transcriptionally up-regulated in the NALT after rmIL-21 stimulation. Moreover, IL-21 signal significantly suppressed production of eotaxin by nasal mucosal fibroblasts, suggesting that IL-21 may interfere with migration of eosinophils. This may be another mechanism underlying the amelioration of local allergic reaction by IL-21⁶⁰.

Conclusion

Recent progress in immunology revealed that the orchestration of the four major CD4⁺ T cell subsets, i.e., Th1, Th2, Th17 and Treg, is pivotal for homeostasis, defense against infection, and pathogenesis and progression of various disorders⁶¹. IL-21 is an essential player that induces and maintains Th17, while physiological and pathological significance of other roles of this pleiotropic cytokine remains to be fully understood. Our studies suggest significant effect of IL-21 signal on IgE-based allergic reaction in vivo, and molecular basis underlying the pathway.

The present data suggest that IL-21 may offer useful therapeutic modalities for allergic disorders. The incidence of allergic disorders is drastically increasing in not only industrial but also developing countries, causing severe medical and social problems. If IgE production can be effectively and safely controlled in patients, such procedures may alleviate allergic symptoms, leading to novel therapeutic and prophylactic interventions of allergic diseases. Finally, molecular analyses on the IL-21-mediated suppression of allergic responses may provide important information that is quite useful for development of novel diagnostic and therapeutic procedures to control allergic disorders.

Acknowledgments

This work was supported by a Grant-in-Aid from the Ministry of Education, Culture, Sports, Science and Technology of Japan.

References

- 1) Parrish-Novak J, Foster DC, Holly RD, Clegg CH. Interleukin-21 and the IL-21 receptor: novel effectors of NK and T cell responses. *J Leukoc Biol* 2002; 72: 856-863.
- 2) Zeng R, Spolski R, Casas E, Zhu W, Levy DE, Leonard WJ. The molecular basis of IL-21-mediated proliferation. *Blood* 2007; 109: 4135-4142.
- 3) Davis ID, Skak K, Smyth MJ, Kristjansen PE, Miller DM, Sivakumar PV. Interleukin-21 signaling: functions in cancer and autoimmunity. *Clin Cancer Res* 2007; 13: 6926-6932.
- 4) Spolski R, Leonard WJ. IL-21 and T follicular helper cells. *Int Immunol* 2010; 22: 7-12.
- 5) Zhou L, Ivanov, II, Spolski R, Min R, Shenderov K, Egawa T, Levy DE, Leonard WJ, Littman DR. IL-6 programs T(H)-17 cell differentiation by promoting sequential engagement of the IL-21 and IL-23 pathways. *Nat Immunol* 2007; 8: 967-974.
- 6) Korn T, Bettelli E, Gao W, Awasthi A, Jager A, Strom TB, Oukka M, Kuchroo VK. IL-21 initiates an alternative pathway to induce proinflammatory T(H) 17 cells. *Nature* 2007; 448: 484-487.
- 7) Nurieva R, Yang XO, Martinez G, Zhang Y, Panopoulos AD, Ma L, Schluns K, Tian Q, Watowich

- SS, Jetten AM, Dong C. Essential autocrine regulation by IL-21 in the generation of inflammatory T cells. *Nature* 2007; 448: 480-483.
- 8) Coquet JM, Kyriakopoulos K, Pellicci DG, Besra G, Berzins SP, Smyth MJ, Godfrey DI. IL-21 is produced by NKT cells and modulates NKT cell activation and cytokine production. *J Immunol* 2007; 178: 2827-2834.
 - 9) Ortega C, Fernandez AS, Carrillo JM, Romero P, Molina IJ, Moreno JC, Santamaria M. IL-17-producing CD8⁺ T lymphocytes from psoriasis skin plaques are cytotoxic effector cells that secrete Th17-related cytokines. *J Leukoc Biol* 2009; 86: 435-443.
 - 10) Parrish-Novak J, Dillon SR, Nelson A, Hammond A, Sprecher C, Gross JA, Johnston J, Madden K, Xu W, West J, Schrader S, Burkhead S, Heipel M, Brandt C, Kuijper JL, Kramer J, Conklin D, Presnell SR, Berry J, Shiota F, Bort S, Hambly K, Mudri S, Clegg C, Moore M, Grant FJ, Lofton-Day C, Gilbert T, Rayond F, Ching A, Yao L, Smith D, Webster P, Whitmore T, Maurer M, Kaushansky K, Holly RD, Foster D. Interleukin 21 and its receptor are involved in NK cell expansion and regulation of lymphocyte function. *Nature* 2000; 408: 57-63.
 - 11) Wurster AL, Rodgers VL, Satoskar AR, Whitters MJ, Young DA, Collins M, Grusby MJ. Interleukin 21 is a T helper (Th) cell 2 cytokine that specifically inhibits the differentiation of naive Th cells into interferon gamma-producing Th1 cells. *J Exp Med* 2002; 196: 969-977.
 - 12) Suto A, Wurster AL, Reiner SL, Grusby MJ. IL-21 inhibits IFN-gamma production in developing Th1 cells through the repression of Eomesodermin expression. *J Immunol* 2006; 177: 3721-3727.
 - 13) Li Y, Bleakley M, Yee C. IL-21 influences the frequency, phenotype, and affinity of the antigen-specific CD8 T cell response. *J Immunol* 2005; 175: 2261-2269.
 - 14) Zeng R, Spolski R, Finkelstein SE, Oh S, Kovanen PE, Hinrichs CS, Pise-Masison CA, Radonovich MF, Brady JN, Restifo NP, Berzofsky JA, Leonard WJ. Synergy of IL-21 and IL-15 in regulating CD8⁺ T cell expansion and function. *J Exp Med* 2005; 201: 139-148.
 - 15) Pelletier M, Bouchard A, Girard D. In vivo and in vitro roles of IL-21 in inflammation. *J Immunol* 2004; 173: 7521-7530.
 - 16) Brandt K, Bulfone-Paus S, Foster DC, Ruckert R. Interleukin-21 inhibits dendritic cell activation and maturation. *Blood* 2003; 102: 4090-4098.
 - 17) Strengell M, Lehtonen A, Matikainen S, Julkunen I. IL-21 enhances SOCS gene expression and inhibits LPS-induced cytokine production in human monocyte-derived dendritic cells. *J Leukoc Biol* 2006; 79: 1279-1285.
 - 18) Jin H, Carrio R, Yu A, Malek TR. Distinct activation signals determine whether IL-21 induces B cell costimulation, growth arrest, or Bim-dependent apoptosis. *J Immunol* 2004; 173: 657-665.
 - 19) Good KL, Bryant VL, Tangye SG. Kinetics of human B cell behavior and amplification of proliferative responses following stimulation with IL-21. *J Immunol* 2006; 177: 5236-5247.
 - 20) Jin H, Malek TR. Redundant and unique regulation of activated mouse B lymphocytes by IL-4 and IL-21. *J Leukoc Biol* 2006; 80: 1416-1423.
 - 21) Mehta DS, Wurster AL, Grusby MJ. Biology of IL-21 and the IL-21 receptor. *Immunol Rev* 2004; 202: 84-95.
 - 22) Ozaki K, Spolski R, Ettinger R, Kim HP, Wang G, Qi CF, Hwu P, Shaffer DJ, Akilesh S, Roopenian DC, Morse HC, 3rd, Lipsky PE, Leonard WJ. Regulation of B cell differentiation and plasma cell generation by IL-21, a novel inducer of Blimp-1 and Bcl-6. *J Immunol* 2004; 173: 5361-5371.
 - 23) Ettinger R, Sims GP, Fairhurst AM, Robbins R, da Silva YS, Spolski R, Leonard WJ, Lipsky PE. IL-21 induces differentiation of human naive and memory B cells into antibody-secreting plasma cells. *J Immunol* 2005; 175: 7867-7879.
 - 24) Mehta DS, Wurster AL, Whitters MJ, Young DA, Collins M, Grusby MJ. IL-21 induces the apoptosis of resting and activated primary B cells. *J Immunol* 2003; 170: 4111-4118.
 - 25) Suto A, Nakajima H, Hirose K, Suzuki K, Kagami S, Seto Y, Hoshimoto A, Saito Y, Foster DC, Iwamoto I. Interleukin 21 prevents antigen-induced IgE production by inhibiting germ line C(epsilon) transcription of IL-4-stimulated B cells. *Blood* 2002; 100: 4565-4573.
 - 26) Ozaki K, Spolski R, Feng CG, Qi CF, Cheng J, Sher A, Morse HC, 3rd, Liu C, Schwartzberg PL, Leonard WJ. A critical role for IL-21 in regulating immunoglobulin production. *Science* 2002; 298: 1630-1634.
 - 27) Rankin AL, MacLeod H, Keegan S, Andreyeva T, Lowe L, Bloom L, Collins M, Nickerson-Nutter C, Young D, Guay H. IL-21 receptor is critical for the development of memory B cell responses. *J Immunol*

- 2011; 186: 667-674.
- 28) di Carlo E, de Toter D, Piazza T, Fabbi M, Ferrini S. Role of IL-21 in immune-regulation and tumor immunotherapy. *Cancer Immunol Immunother* 2007; 56: 1323-1334.
- 29) Kishida T, Asada H, Itokawa Y, Cui FD, Shin-Ya M, Gojo S, Yasutomi K, Ueda Y, Yamagishi H, Imanishi J, Mazda O. Interleukin (IL)-21 and IL-15 genetic transfer synergistically augments therapeutic antitumor immunity and promotes regression of metastatic lymphoma. *Mol Ther* 2003; 8: 552-558.
- 30) Nakano H, Kishida T, Asada H, Shin-Ya M, Shinomiya T, Imanishi J, Shimada T, Nakai S, Takeuchi M, Hisa Y, Mazda O. Interleukin-21 triggers both cellular and humoral immune responses leading to therapeutic antitumor effects against head and neck squamous cell carcinoma. *J Gene Med* 2006; 8: 90-99.
- 31) Spolski R, Leonard WJ. Interleukin-21: Basic Biology and Implications for Cancer and Autoimmunity. *Annu Rev Immunol* 2007; 26: 57-59.
- 32) Li J, Pan HF, Cen H, Tian J, Ma Y, Tao JH, Ye DQ. Interleukin-21 as a potential therapeutic target for systemic lupus erythematosus. *Mol Biol Rep* 2010; (published on line).
- 33) Costanzo A, Chimenti MS, Botti E, Caruso R, Sarra M, Monteleone G. IL-21 in the pathogenesis and treatment of skin diseases. *J Dermatol Sci* 2010; 60: 61-66.
- 34) Barker BR, Gladstone MN, Gillard GO, Panas MW, Letvin NL. Critical role for IL-21 in both primary and memory anti-viral CD8(+) T-cell responses. *Eur J Immunol* 2010; 40: 3085-3096.
- 35) Hecker M, Bohnert A, Konig IR, Bein G, Hackstein H. Novel genetic variation of human interleukin-21 receptor is associated with elevated IgE levels in females. *Genes Immun* 2003; 4: 228-233.
- 36) Chatterjee R, Batra J, Ghosh B. A common exonic variant of interleukin21 confers susceptibility to atopic asthma. *Int Arch Allergy Immunol* 2009; 148: 137-146.
- 37) Shang XZ, Ma KY, Radewonuk J, Li J, Song XY, Griswold DE, Emmell E, Li L. IgE isotype switch and IgE production are enhanced in IL-21-deficient but not IFN-gamma-deficient mice in a Th2-biased response. *Cell Immunol* 2006; 241: 66-74.
- 38) Wood N, Bourque K, Donaldson DD, Collins M, Vercelli D, Goldman SJ, Kasaian MT. IL-21 effects on human IgE production in response to IL-4 or IL-13. *Cell Immunol* 2004; 231: 133-145.
- 39) Caven TH, Shelburne A, Sato J, Chan-Li Y, Becker S, Conrad DH. IL-21 dependent IgE production in human and mouse in vitro culture systems is cell density and cell division dependent and is augmented by IL-10. *Cell Immunol* 2005; 238: 123-134.
- 40) Pene J, Guglielmi L, Gauchat JF, Harrer N, Woisetschlager M, Boulay V, Fabre JM, Demoly P, Yssel H. IFN-gamma-mediated inhibition of human IgE synthesis by IL-21 is associated with a polymorphism in the IL-21R gene. *J Immunol* 2006; 177: 5006-5013.
- 41) Kishida T, Hiromura Y, Shin-Ya M, Asada H, Kuriyama H, Sugai M, Shimizu A, Yokota Y, Hama T, Imanishi J, Hisa Y, Mazda O. IL-21 Induces Inhibitor of Differentiation 2 and Leads to Complete Abrogation of Anaphylaxis in Mice. *J Immunol* 2007; 179: 8554-8561.
- 42) Honjo T, Kinoshita K, Muramatsu M. Molecular mechanism of class switch recombination: linkage with somatic hypermutation. *Annu Rev Immunol* 2002; 20: 165-196.
- 43) Muramatsu M, Kinoshita K, Fagarasan S, Yamada S, Shinkai Y, Honjo T. Class switch recombination and hypermutation require activation-induced cytidine deaminase (AID), a potential RNA editing enzyme. *Cell* 2000; 102: 553-563.
- 44) Honjo T, Nagaoka H, Shinkura R, Muramatsu M. AID to overcome the limitations of genomic information. *Nat Immunol* 2005; 6: 655-661.
- 45) Longeri S, Basu U, Alt F, Storb U. AID in somatic hypermutation and class switch recombination. *Curr Opin Immunol* 2006; 18: 164-174.
- 46) Qiu G, Stavnezer J. Overexpression of BSAP/Pax-5 inhibits switching to IgA and enhances switching to IgE in the L.29 mu B cell line. *J Immunol* 1998; 161: 2906-2918.
- 47) Sayegh CE, Quong MW, Agata Y, Murre C. E-proteins directly regulate expression of activation-induced deaminase in mature B cells. *Nat Immunol* 2003; 4: 586-593.
- 48) Fenghao X, Saxon A, Nguyen A, Ke Z, Diaz-Sanchez D, Nel A. Interleukin 4 activates a signal transducer and activator of transcription (Stat) protein which interacts with an interferon-gamma activation site-like sequence upstream of the I epsilon exon in a human B cell line. Evidence for the involvement of Janus kinase 3 and interleukin-4 Stat. *J Clin Invest* 1995; 96:

- 907-914.
- 49) Dedeoglu F, Horwitz B, Chaudhuri J, Alt FW, Geha RS. Induction of activation-induced cytidine deaminase gene expression by IL-4 and CD40 ligation is dependent on STAT6 and NFkappaB. *Int Immunol* 2004; 16: 395-404.
- 50) Thienes CP, De Monte L, Monticelli S, Busslinger M, Gould HJ, Vercelli D. The transcription factor B cell-specific activator protein (BSAP) enhances both IL-4- and CD40-mediated activation of the human epsilon germline promoter. *J Immunol* 1997; 158: 5874-5882.
- 51) Quong MW, Harris DP, Swain SL, Murre C. E2A activity is induced during B-cell activation to promote immunoglobulin class switch recombination. *Embo J* 1999; 18: 6307-6318.
- 52) Takeda K, Tanaka T, Shi W, Matsumoto M, Minami M, Kashiwamura S, Nakanishi K, Yoshida N, Kishimoto T, Akira S. Essential role of Stat6 in IL-4 signalling. *Nature* 1996; 380: 627-630.
- 53) Yokota Y. Id and development. *Oncogene* 2001; 20: 8290-8298.
- 54) Sugai M, Gonda H, Nambu Y, Yokota Y, Shimizu A. Role of Id proteins in B lymphocyte activation: new insights from knockout mouse studies. *J Mol Med* 2004; 82: 592-599.
- 55) Murre C. Helix-loop-helix proteins and lymphocyte development. *Nat Immunol* 2005; 6: 1079-1086.
- 56) Lasorella A, Uo T, Iavarone A. Id proteins at the cross-road of development and cancer. *Oncogene* 2001; 20: 8326-8333.
- 57) Ruzinova MB, Benezra R. Id proteins in development, cell cycle and cancer. *Trends Cell Biol* 2003; 13: 410-418.
- 58) Gonda H, Sugai M, Nambu Y, Katakai T, Agata Y, Mori KJ, Yokota Y, Shimizu A. The balance between Pax5 and Id2 activities is the key to AID gene expression. *J Exp Med* 2003; 198: 1427-1437.
- 59) Sugai M, Gonda H, Kusunoki T, Katakai T, Yokota Y, Shimizu A. Essential role of Id2 in negative regulation of IgE class switching. *Nat Immunol* 2003; 4: 25-30.
- 60) Hiromura Y, Kishida T, Nakano H, Hama T, Imanishi J, Hisa Y, Mazda O. IL-21 administration into the nostril alleviates murine allergic rhinitis. *J Immunol* 2007; 179: 7157-7165.
- 61) Korn T, Bettelli E, Oukka M, Kuchroo VK. IL-17 and Th17 Cells. *Annu Rev Immunol* 2009; 27: 485-517.

〈和文抄録〉

IL-21 シグナルとアレルギー免疫応答

松田 修, 岸田 綱郎

京都府立医科大学大学院医学研究科免疫・微生物学

IL-21 は、 γ 鎖サイトカインファミリーに属し、主として Th17 と NKT 細胞から産生される。T, B, NK 細胞, マクロファージ, 樹状細胞等に作用して多彩な免疫応答制御を司る。IL-21 はまた、自己免疫疾患や悪性腫瘍等の病態にも関与する。我々は、IL-21 シグナルが B 細胞に Id2 分子の発現を誘導し、この Id2 が必須に関与することで、IL-4 シグナルに拮抗して IgE クラススイッチを抑制することを見出した。さらに、アナフィラキシーや局所アレルギー病態を *in vivo* で抑制することも示した。IL-21 によるアレルギー抑制機構を分子レベルで理解することは、獲得免疫応答のネットワーク制御における IL-21 の意義を理解することにつながるのみならず、アレルギー疾患の新しい治療手段の開発につながる可能性がある。

キーワード：インターロイキン-21, Id2, IgE, クラススイッチ, アレルギー。

著者プロフィール



松田 修 Osam Mazda

所属・職：京都府立医科大学大学院医学研究科免疫・微生物学・教授

略 歴：1985年3月 京都府立医科大学医学部医学科 卒業

1985年4月 自治医科大学附属病院内科

1993年3月 京都大学大学院医学研究科博士課程修了 医学博士

1993年4月 日本学術振興会特別研究員

1994年10月 京都府立医科大学微生物学教室助手

1995年4月 同講師

1999年6月 同助教授

2008年6月～ 日本学術振興事業団さきがけ研究員（兼務）

2010年6月～ 現職

専門分野：免疫学，再生医学

主な業績：1. Hiromura, Y. et al. J Immunol. 179: 7157-7165, 2007.

2. Kishida T. et al. J Immunol. 179: 8554-8561, 2007.

3. Matsui M, et al. Cancer Res. 69: 2523-30. 2009.

著者プロフィール



岸田 綱郎 Tsunao Kishida

所属・職：京都府立医科大学大学院医学研究科免疫・微生物学・学内講師

略 歴：1997年3月 明治国際医療大学鍼灸学部 卒業

1999年4月 明治国際医療大学大学院博士前期課程修了

2003年3月 京都府立医科大学大学院医学研究科博士課程修了

2003年4月 (財)ルイ・パストゥール医学研究センター研究員

2005年7月 (財)ルイ・パストゥール医学研究センター研究室長

2005年7月 (財)ルイ・パストゥール医学研究センター理事

2006年10月 京都府立医科大学博士研究員

2010年7月 京都府立医科大学大学院医学研究科免疫・微生物学助教

2011年1月～ 現職

専門分野：免疫学，再生医学

主な業績：1. Hiromura, Y. et al. J Immunol. 179: 7157-7165, 2007.

2. Kishida T. et al. J Immunol. 179: 8554-8561, 2007.

3. Matsui M, et al. Cancer Res. 69: 2523-30. 2009.